# The cDNA of the two isoforms of bovine cGMP-dependent protein kinase

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cDNAs encoding the isoform I $\alpha$  of the cGMP-dependent protein kinase were isolated from a bovine trachea smooth muscle cDNA library constructed in  $\lambda$ gt10. The deduced protein sequence is identical with the protein sequence obtained by Edman degradation of the bovine lung enzyme [(1984) Biochemistry 23, 4207–4218]. Alternate cDNA clones were isolated which code for a protein slightly different within the aminoterminal part from the known amino acid sequence. These alternate cDNAs contain the sequence of a peptide identified in the isoform I $\beta$  of cGMP-dependent protein kinase. Northern blot analysis of poly(A)<sup>+</sup> RNA from bovine trachea smooth muscle indicated the presence of two different mRNA species of about 6.2 kb.

Protein kinase, cyclic GMP-dependent; cDNA cloning; Smooth muscle

## 1. INTRODUCTION

Cyclic GMP-dependent protein kinase (cGMPkinase) is one of the main intracellular receptors for cGMP [1]. The native enzyme consists of two identical 76331 Da subunits [2] and is activated by the binding of 4 mol cGMP per mol holoenzyme [3,4]. Each subunit of cGMP-kinase contains a regulatory part – the aminoterminus including the autophosphorylation site and two distinct cGMP binding sites - and a catalytic part: the ATP binding site and the catalytic center. Sequence analysis of various protein kinases and cyclic nucleotide binding proteins has led to the hypothesis that cGMP-kinase is the product of a fusion between two ancestral genes which code for a cyclic nucleotide binding protein and a protein kinase [2].

Cyclic GMP-kinase is present in high concentrations in smooth muscle [5-8]. The active enzyme decreases cytosolic levels in tracheal [7] and vascular [8] smooth muscle by an unidentified

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mechanism and thereby induces relaxation. Two isoforms (I $\alpha$  and I $\beta$ ) which apparently are functionally identical but may differ in the expression of their mRNA have been identified in bovine aorta [9,10]. To facilitate further studies on the physiological role of cGMP-kinase, the cDNA of cGMP-kinase mRNA has been cloned from bovine trachea smooth muscle.

## 2. MATERIALS AND METHODS

## 2.1. Construction and screening of cDNA libraries

Poly(A)<sup>+</sup> RNA was isolated from bovine trachea smooth muscle according to [11], followed by oligo(dT)-cellulose column chromatography [12]. Specific and random primed cDNA libraries were constructed in  $\lambda gt10$  according to [13]. 2  $\mu g$  poly(A)<sup>+</sup> RNA yielded 5  $\times$  10<sup>6</sup> independent recombinant phages.

The libraries were screened without prior amplification by synthetic oligonucleotide probe mixtures (fig.1, inset) and isolated cDNA fragments. The 5' ends of the oligonucleotides were labeled using T<sub>4</sub> polynucleotide kinase. Isolated cDNA fragments were labeled by random priming [14].

# 2.2. Subcloning and sequencing of cDNA inserts

Positive clones were plaque purified, their cDNA inserts isolated and subcloned into the *EcoRI* or *SmaI* site of pUC18. Plasmid DNA suitable for sequencing was prepared by the alkaline lysis method [15]. Sequence analysis was performed by

the dideoxy chain termination method [16] using  $[\alpha^{-35}S]dATP$  and modified T7 DNA-polymerase (USB). Most of the sequence was obtained by sequencing both strands of the cDNA independently.

#### 2.3. Northern blot analysis

Poly(A)<sup>+</sup> RNA was denatured with glyoxal (1 M) and 50% dimethylsulfoxide, electrophoresed on 1.5% agarose gels and transferred to Biodyne Nylon membranes [17]. Blot hybridization was carried out with cDNA fragments labeled by the random priming method [14]. If not stated otherwise all other cloning procedures were carried out according to [17].

## 3. RESULTS

# 3.1. Isolation of cDNA clones for isoform $I\alpha$

The cloning strategy is summarized in fig.1. Twelve 14-mer oligodeoxynucleotides complementary to all possible codons for the carboxyterminal amino acid sequence Trp-Asp-Ile-Asp-Phe (excluding the third nucleotide residue of the Phecodon) of cGMP-kinase were synthesized and used for construction of a primer extension library. About  $5 \times 10^5$  recombinant phages were screened with two different probes. One probe (A) contained an equimolar mixture of 64 oligonucleotides (21-mer) representing all the possible cDNA sequences corresponding to amino acid 618-624

(fig.1, amino acid numbers (fig.2) correspond to those in [2] + 1 since the sequence of [2] lacks the initiation amino acid Met). The second probe (B) (42 bp, aa 648-661) was designed according to codon usage [18]. The clone λcGpeI-6.2 was isolated, subcloned and sequenced. The cDNA sequence (215 bp, nt 1799-2014 of the complete sequence, fig.2) contained the sequence of the primer and of both probes. The insert of this clone was used to rescreen the same library and to isolate clone  $\lambda$ cGpeI-8.2. Sequencing of this insert (nt 601-2010, fig.2) showed that it contained an open reading frame coding for an amino acid sequence corresponding to residues 200 (Ile) to 670 (Asp) of cGMP-kinase [2]. Using the 5'-terminal HindIII (nt 695)-HindIII (nt 1316) fragment one clone  $\lambda$ cGrp-5.1 was isolated by screening approx. 1  $\times$ 10<sup>6</sup> independent clones of a random primed cDNA library. The cDNA insert of this clone (1810 bp) contained the sequence from nucleotide 959 to 2769 (fig.2).

A synthetic primer complementary to nt 828-846 (fig.2) was used to construct a second primer extension library (fig.1). One million recombinant phages of this library were screened using the nt 601-695 fragment as probe. Eleven

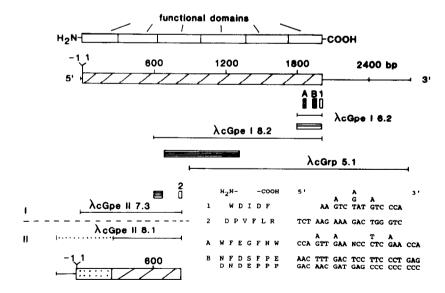


Fig. 1. Cloning strategy. (Top) Functional domains of the amino acid sequence of cGMP-kinase according to [2]. Crossed bar and solid line represent the protein coding and the 5'- and 3'-noncoding region of the nucleotide sequence, respectively. (Bottom) The length of the cDNA inserts of individual clones are shown by lines. The open and hatched bars indicate the position of primers 1 and 2 and the screening probes, respectively. (I) Isoform  $I\alpha$ ; (II) isoform  $I\alpha$ ; the dotted part of  $I\alpha$  represents that part which is not identical with  $I\alpha$ . (Inset) Amino acid sequences and derived synthesized nucleotide sequences of primers 1 and 2 and screening probes A and B.

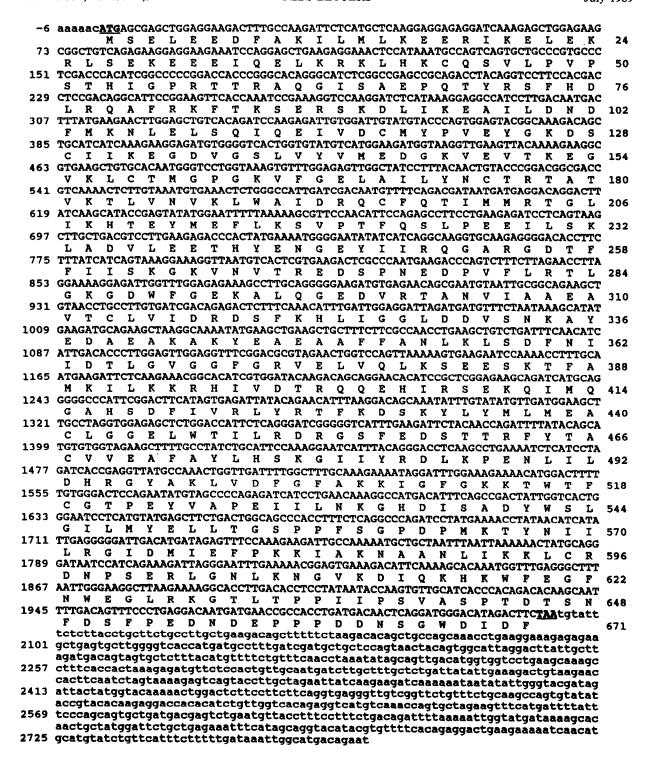


Fig.2. Nucleotide and deduced amino acid sequences of isoform  $I\alpha$  of cGMP-kinase. The translational initiation signal ATG and the termination signal TAA are underlined.

positive clones including clone  $\lambda$ cGpeII-7.3 were isolated. This clone contained the cDNA sequence from nucleotide -6 to 827 (numeration starts with +1 at the A of the initiation codon ATG) of cGMP-kinase.

Fig.2 shows the composed nucleotide sequence and the deduced amino acid sequence of the cDNA of isoform  $I\alpha$  of the cGMP-kinase from bovine trachea smooth muscle. The composed nucleotide sequence consisted of 2775 bp and contained an open reading frame of 2013 bp coding for a protein with a calculated molecular mass of 76418 Da and also part of the 5'- and 3'-noncoding sequence. The deduced amino acid sequence is identical to the sequence of the bovine enzyme [2] with the addition of the initiation Met.

# 3.2. Isolation of cDNA clones for isoform IB

In addition to  $\lambda$ cGpeII-7.3 the clone  $\lambda$ cGpeII-8.1 was isolated from the second primer extension library (fig.1). The cDNA insert of this clone (1042 nt, fig.3) contained an open reading frame of 879 bp including the initiation codon ATG (A = +1 in the numeration) and 163 bp of the 5'-noncoding region of the isoform  $I\beta$  mRNA. From nucleotide 312 to the 3'-end of the clone (nt 879) the nucleotide and the deduced amino acid se-

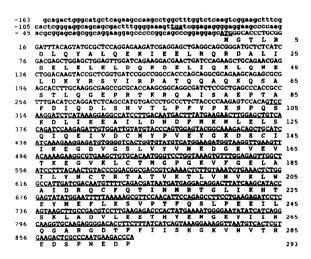


Fig.3. Nucleotide and deduced amino acid sequences of the aminoterminal part of isoform  $I\beta$  of cGMP-kinase. The nucleotide sequence of  $\lambda$ cGpeII-8.1 which is identical to the isoform  $I\alpha$ -sequence is underlined (Ser<sup>105</sup> of isoform  $I\beta$  = Ser<sup>90</sup> of isoform  $I\alpha$ ). The initiation codon ATG and the upstream termination codon TGA are also underlined.

quence were identical to the corresponding part of the isoform  $I\alpha$  (nt 267 to nt 834). To confirm the second isoform the sequence nt 312-879 of clone  $\lambda$ cGpeII-8.1, which is present in both isoforms, was used to rescreen the same library. Ten clones were isolated and sequenced. Seven and 3 clones coded for isoform  $I\alpha$  and  $I\beta$ , respectively.

### 3.3. Northern blot analysis

The 3'-noncoding and the protein coding sequence of isoform  $I\alpha$  were used for Northern blot analysis of poly(A)+ RNA from bovine trachea smooth muscle (fig.4). These probes hybridized to a mRNA species of approx. 6.2 kb suggesting that the mRNA for cGMP-kinase contains approx. 4.2 kb noncoding sequences. The 5'-noncoding and that part of the coding sequence of  $\lambda$ cGpeII-8.1 which was not identical to isoform I $\alpha$ also hybridized to a 6.2 kb mRNA species (fig.4). In agreement with the isolation frequency of the cDNA clones the message for the isoform IB was consistently less (approx. 10%) than that of isoform I $\alpha$  (fig.4) suggesting that each isoform of cGMP-kinase is coded by a distinct mRNA species of approximately identical size.

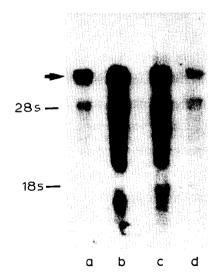


Fig. 4. Northern blot analysis of RNA from bovine trachea smooth muscle. (a,b) 5 and 20  $\mu$ g of poly(A)<sup>+</sup> RNA were hybridized with a probe for isoform  $I\alpha$  (nt 959–2769). Identical results were obtained when the sequence nt 695–1316 ( $I\alpha$ ) was used as a probe (not shown); (c,d) 20 and 5  $\mu$ g of poly(A)<sup>+</sup> RNA were hybridized with a probe specific only for isoform  $I\beta$  ( $\lambda$ cGpeII-8.1, nt – 163 to 312). The probes were labeled to the same specific activity.

## 4. DISCUSSION

Two distinct cDNAs for cGMP-kinase were identified. The derived amino acid sequence for isoform  $I\alpha$  is identical with the known sequence of bovine lung cGMP-kinase [2]. The second derived amino acid sequence ( $I\beta$ ) had an extended

aminoterminus which differs from that of isoform  $I\alpha$ . No evidence was obtained for any differences between the two isoforms in the remaining part of the coding region which contains the two cGMP-and the ATP-binding sites and the catalytic center. The in vivo occurrence and identity of isoform  $I\beta$  is further supported by (i) the identity of aa 66-75

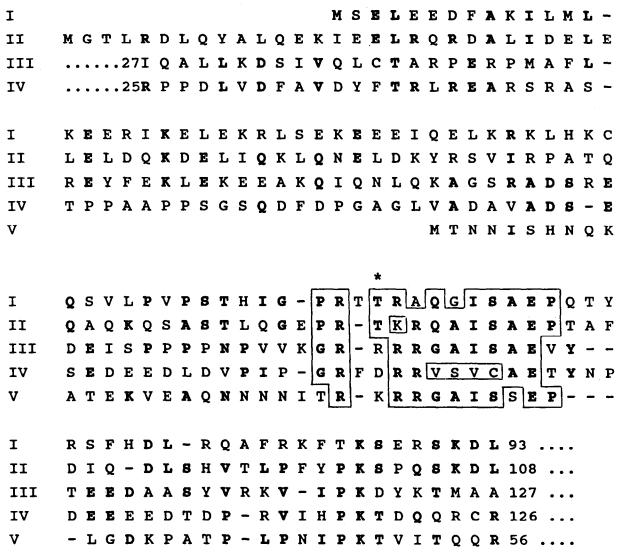


Fig. 5. Comparison of the aminoterminal sequences of cGMP-kinases and regulatory subunits of cAMP-kinase. Amino acid sequence alignment of part of the regulatory domains of isoforms  $I\alpha$  and  $I\beta$  of cGMP-kinase and of part of the regulatory subunits of different types of the cAMP-kinase. The sequences were aligned by eye. Gaps are marked with a dash. Identical amino acids are shown in bold. The hinge region containing the autophosphorylation sites of cGMP-kinase  $I\alpha$  (\* = autophosphorylated Thr) and RII $\alpha$  is boxed. (I) aa 1-93 of isoform  $I\alpha$  of cGMP-kinase; (II) aa 1-108 of isoform  $I\beta$  of cGMP-kinase; (III) aa 27-127 of R $I\alpha$  of cAMP-kinase (bovine skeletal muscle) [26]; (IV) aa 25-126 of RII $\alpha$  of cAMP-kinase (bovine heart) [27]; (V) aa 1-56 of R of cAMP-kinase (Dictyostelium discoideum) [28].

of the derived amino acid sequence with a peptide sequence published for isozyme I $\beta$  [10], and (ii) the identification of two distinct mRNA species for isoforms I $\alpha$  and I $\beta$ . However, the concentration of the mRNA for isoform I $\beta$  is about 10% of that of isoform I $\alpha$ . The mRNA for both isoforms exceeds the coding region about 2-fold. This is not unusual since similar large mRNA species were described for the regulatory subunits RI (2.4-4.3 kb) [20-22] and RII (6.0 kb) [23,24] of cAMP-kinase.

The two isoforms from bovine trachea smooth muscle are very similar. Using the Dayhoff MDM-78 comparison matrix [25], the aminoterminus (aa 1-108) of the deduced amino acid sequence of isoform LB shows a high degree of similarity to isoform I $\alpha$  (71.2%, as 1-93) and to the corresponding parts of the regulatory subunits of cAMP-kinase (RI $\alpha$  from bovine skeletal muscle [26] 64.5% (aa 27–127); RII $\alpha$  from bovine heart [27] 62.0% (aa 25-126) and R from Dictvostelium discoideum [28] 63.3% (aa 1-56)) (fig.5). The highest degree of similarity is found in the socalled 'hinge region' [29] which contains at Thr<sup>59</sup> the autophosphorylation site of cGMP-kinase  $I\alpha$ [2,30]. The difference in the aminoterminus of isoforms I $\alpha$  and I $\beta$  is very interesting since it has been shown that the aminoterminus of I $\alpha$  is that part of the enzyme which regulates binding of cGMP to and activation of the cGMP-kinase [4].

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